School of Medical and Allied Sciences

Course Code: BPHT 3003 Course Name: Pharmaceutical Microbiology

Microbiological Assay Types and Technique

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Disclaimer

All the content material provided here is only for teaching purpose.



Introduction

- A microbiological assay defined as qualitative or quantitative determination of chemical compound from a simple or even complex material with the use of microorganisms.
- Also called **microbial assay**
- Used to determine the potency of a drug in animals or man and monitoring and controlling antimicrobial chemotherapy
- Many anti-microbial agents, which inhibit the growth of microorganisms (antibiotics) or are essential for their growth (vitamins and amino acids).
- RELATIVE POTENCY: defined as the term used to express the biological activity of a sample preparation compared to a standard preparation.

Advantages:

- **Simple**
- **Specific**
- Minimum requirements of space, labour, materials and time
- Inexpensive and
- **Convenient method Disadvantages:**
- Required skilled technician
- Required proper calibration
- **Less reproducible**
- **Chances of greater error**
- **❖** Not used if a good alternative physical or chemical assay is available

Microbiological assay of an antibiotic

- The microbiological assay of an antibiotic is based upon a comparison of the inhibition of growth of micro-organisms by measured concentrations of the antibiotics under examination with that produced by known concentrations of a standard preparation of the antibiotic having a known activity.
- To be utilised for demonstrating the therapeutic efficacy of antibiotics.
- Assay: it is a physical or biological testing method to determine content or quality of substances.

Two general methods are usually employed:

- I) The cylinder-plate (or cup-plate) method (method A) and
- II) the turbidimetric (or tube assay) method(method B)
- The cylinder-plate method depends upon diffusion of the antibiotic from a vertical cylinder through a solidified agar layer in a Petri dish or plate to an extent such that growth of the added micro-organism is prevented entirely in a zone around the cylinder containing a solution of the antibiotic.
- The turbidimetric method depends upon the inhibition of growth of a microbial culture in a uniform solution of the antibiotic in a fluid medium that is favourable to its rapid growth in the absence of the antibiotic.

Culture media used for antibiotic assay

- Dissolved all the required ingredients (table1) in sufficient water to produced 1000ml
- pH can be maintained by adding 1M solution of HCl or NaOH.
- Followed by the sterilization of the media
- FTM, ATM and SCDM are used.

Composition of FTM, ATM and SCDM????? Table 13.1: Composition of media: Quantities in gm per 1000 ml medium

| | | | | | | | S | | | |
|-----------------------------|------|------|-----|------|------|------|------|------|------------|-------|
| Ingredient | A | В | С | D | E | F | G | Н | 1 | J |
| Peptone | 6.0 | 6.0 | 5.0 | 6.0 | 6.0 | 6.0 | 9.4 | - | 10.0 | , - N |
| Yeast extract | 3.0 | 3.0 | 1.5 | 3.0 | 3.0 | 3.0 | 4.7 | - | - | - |
| Beef extract | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 2.4 | • | 10.0 | _ |
| Pancreatic digest of casein | 4.0 | - | | 4.0 | - | = | | 17.0 | - | 15.0 |
| Dextrose | 1.0 | - | 1.0 | 1.0 | | - | 10.0 | 2.5 | a - | - |
| Papaic digest of soyabean | - | - | _ | - | - | | - | 3.0 | - | 5.0 |
| Agar | 15.0 | 15.0 | - | 15.0 | 15.0 | 15.0 | 23.5 | 12.0 | 17.0 | 15.0 |
| Glycerin | | - | | | - | _ | - | : | 10.0 | 1.50 |

3.5

3.68

1.32

6.95 -

7.05

7.8 -

8.0

7.8 -

8.0

5.8

-6.0

6.5

-6.6

6.5

-6.6

10.0

5.0

2.5

7.1

-7.3

10.0

6.0

-6.2

5.0

7.2

-7.4

3.0

6.9

-7.1

| Pancreatic digest of casein | 4.0 | _ | - | 4.0 | - | | r | 17.0 | - |
|-----------------------------|-----|---|-----|-----|---|----|---------------|------|------|
| Dextrose | 1.0 | - | 1.0 | 1.0 | - | 1- | 10.0 | 2.5 | - a- |
| Papaic digest of soyabean | - | - | - | - | _ | _ | 0 | 3.0 | |

Polysorbate 80

Sodium chloride

Final pH

Dipotassium hydrogen phosphate

Potassium dihydrogen phosphate

Buffer solution

- Prepare by dissolving the following quantities given in Table 2.
- sufficient water to produce 1000 ml after sterilisation, adjusting the pH with 8 M phosphoric acid or 10 M potassium hydroxide.

| Buffer No. | Dipotassium Hydrogen Phosphate, K ₂ HPO ₄ (g) | Potassium Dihydrogen phosphate, KH ₂ PO ₄ (g) | pH adjusted after sterilization to |
|---------------|--|--|--|
| 1 | 2.0 | 8.0 | 6.0±0.1 |
| 2 | 16.73 | 0.523 | 8.0±0.1 |
| 3 | _ | 13.61 | 4.5±0.1 |
| 4 | 20.0 | 80.00 | 6.0±0.1 |
| 5 | 35.0 | _ | 10.5±0.1* |
| 6 | 13.6 | 4.0 | 7.0±0.2 |

Preparation of standard

- A Standard Preparation is an authentic sample of the appropriate antibiotic for which the potency has been precisely determined by reference to the appropriate international standard.
- The Potency of the standard preparation may be expressed in International Units or in µg per mg of the pure antibiotic.
- Ex: Dissolve a quantity of the standard preparation of a given antibiotics in the solvents(table3). Dilute the preparation to get the required concentration as stated and stored in a refrigerator.
- Usually prepared in the ratio of 1:1.5

| Table 13. | 2: Stock solu | tions and test d | | | 1 |
|-----------------------|----------------|--|---------------------------------|--|---|
| | method | Initial solvent for std. stock solution | Final std. stock conc"/mi | Final diluent for test dilution | Median dose ug or Units/ml of test sol*. |
| Amikacin | В | Water | 1 mg | Water | 10 µg |
| Amphotericin B | ^ | Dimethyl sulphoxide | 1 mg | 85 | 1 μg |
| Bacitracin | A | 0.01 M HCI | 100 units | B1 | 1 unit |
| Bleomycin | ^ | B6 | 2 units | B6 | 0.04 unit |
| Capreomycin | В | Water | 1 mg | Water | 100 µg |
| Carbenicillin | A | B1 | 1 mg | B6 | 20 µg |
| Chlortetracycline | A¹ | 0.1 M HCI | 1 mg | Water | 2.5 µg |
| (II | B ² | 0.1 M HCI | 1 mg | Water | 0.24 µg |
| Colistimethate sodium | ^ | Water | 1 mg | 84 | 1 unit |
| u i | 8 | Water | 1 mg | B6 | 1 unit |
| Colistin sulphate | _ | Water | 1 mg | B6 | 1 μg |
| Erythromycin | _ | Methanol | 1 mg | B2 | 1 μg |
| Framycetin | _ | B2 | 1 mg | B2 | 1 μg |
| Gentamicin | A | B2 | 1 mg | B2 | 0.1 µg |
| Kanamycin Sulphate | A¹ | B2 | 800 units | B2 | 0.8 unit |
| | в³ | Water | 1000 units | Water | 10 unit |
| Neomycin | A | B2 | 1 mg | 82 | 1 μg |
| Novobiocin | A | Ethanol | 1 mg | B4 | 0.5 μg |
| Nystatin | A | Dimethyl formamide | 1000 units | B4 | 20 unit |
| Onterraciona | A* | 0.1 M HCI | 1 mg | B3 | 2.5 µg |
| Oxytetracycline | B, | 0.1 M HCI | 1 mg | Water | 0.24 µg |
| Prot. | | Water | 10,000 units | 84 | 10 unit |
| Folymyxin B | A3 | Methanol | 1 mg | B2 | 12-50 units |
| Spramycin | | Water | 1 mg | Water | 1 μg |
| S'eptomycin | A3 | Water | 1 mg | Water | 30 µg |
| | B° | 0.1 M HCI | 1 mg | Water | 2.5 μg |
| etracycline | A ⁴ | O.1 M HCI | 1 mg | Water | 0.24 µg |
| | в' | | 1 mg | Water | 2.5 μg |
| Tobramycin | В | Water | 1 mg | B3 | 10 μg |
| Vancomycin | A | Water | | | P.T.O |

Preparation of the test sample

- From the information available for the substance under examination (the "unknown"), assign to it an assumed potency per unit weight or volume, and on this assumption prepare on the day of the assay a stock solution and test dilution as specified for each antibiotic in Table4 but with the same final diluents as used for the Standard Preparation.
- The assay with 5 levels of the Standard requires only one level of the unknown at a concentration assumed equal to the median level of the standard.

Preparation of Test organism

• The test organism for each antibiotic is listed in Table, together with its identification number in the American Type Culture Collection (ATCC).

| Antibiotic | Test Organism | ATCC1 No. |
|--------------------|-------------------------------|-----------|
| Amikacin | Staphylococcus aureus | 29737 |
| Amphotericin B | Saccharomyces cerevisiae | 9763 |
| Bacitracin | Micrococcus luteus | 10240 |
| Bleomycin | Mycobacterium smegmatis | 607 |
| Carbenicillin | Pseudomonas aeruginosa | 25619 |
| Chlortetracycline | Bacillus pumilus | 14884 |
| Erythromycin | Micrococcus luteus | 9341 |
| Framycetin | Bacillus pumilus | 14884 |
| | Bacillus subtilis | 6633 |
| Gentamicin | Staphylococcus epidermidis | 12228 |
| Kanamycin sulphate | Bacillus pumilus | 14884 |
| | Staphylococcus aureus | 29737 |
| Neomycin | Staphylococcus epidermidis | 12228 |
| Novobiocin | Staphylococcus epidermidis | 12228 |
| Nystatin | Saccharomyces cerevisiae | 2601 |
| Oxytetracycline | Bacillus cereus var, mycoides | 11778 |
| | Staphylococcus aureus | 29737 |
| Polymyxin B | Bordetella bronchiseptica | 4617 |
| Spiramycin | Bacillus pumilus | 6633 |
| Streptomycin | Bacillus subtilis | 6633 |
| | Klebsiella pnumoniae | 10031 |
| Tetracycline | Bacillus cereus | 11778 |
| | Staphylococcus aureus | 29737 |
| Tobramycin | Staphylococcus aureus | 29737 |
| Tylosin | Staphylococcus aureus | 9144 |

Preparation of inoculums

• Inoculums is the mixture of microbes along with the culture media in which it is growing.

Steps involved:

- Maintain the test microbes on slant of medium A and transfer to a fresh slant once a week.
- Incubate the slant at the specified temperature for 1day
- ❖ Using 3ml of slant solution, wash the microbes from agar slant on to a large surface of medium A such as a Roux bottle containing 250ml of agar media
- ❖ Incubate for 1day at the required temperature
- ❖ Wash the growth from the nutrient surface using 50ml of saline solution.
- Store the test microbes under refrigerator



Methods of Microbiological Assay

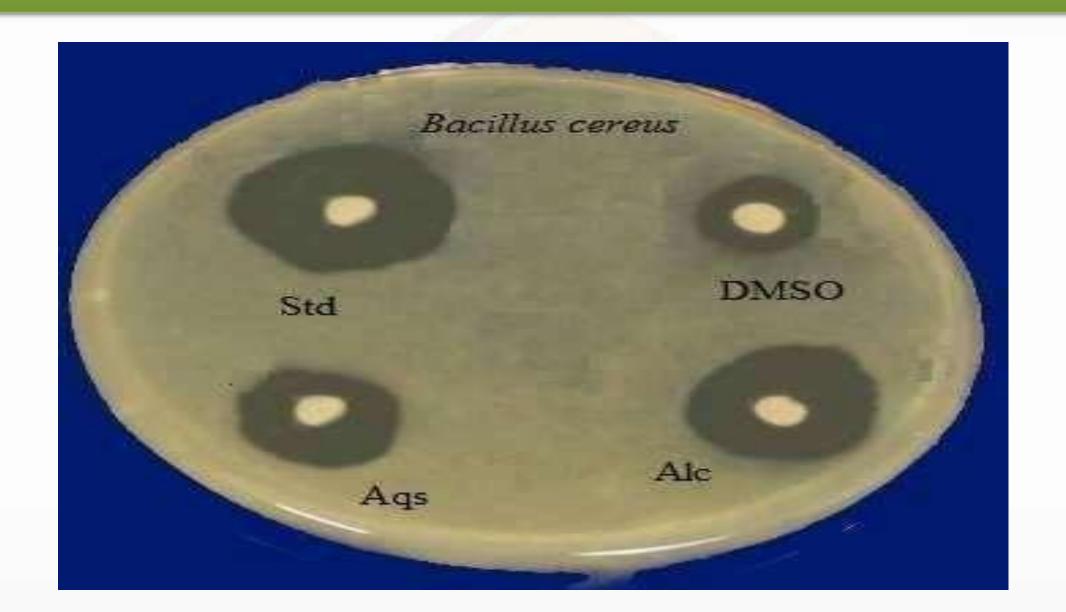
- A. Cylinder plate or Cup plate method
- B. Turbidimetric or Tube Assay method

A. Cylinder plate or cup plate method

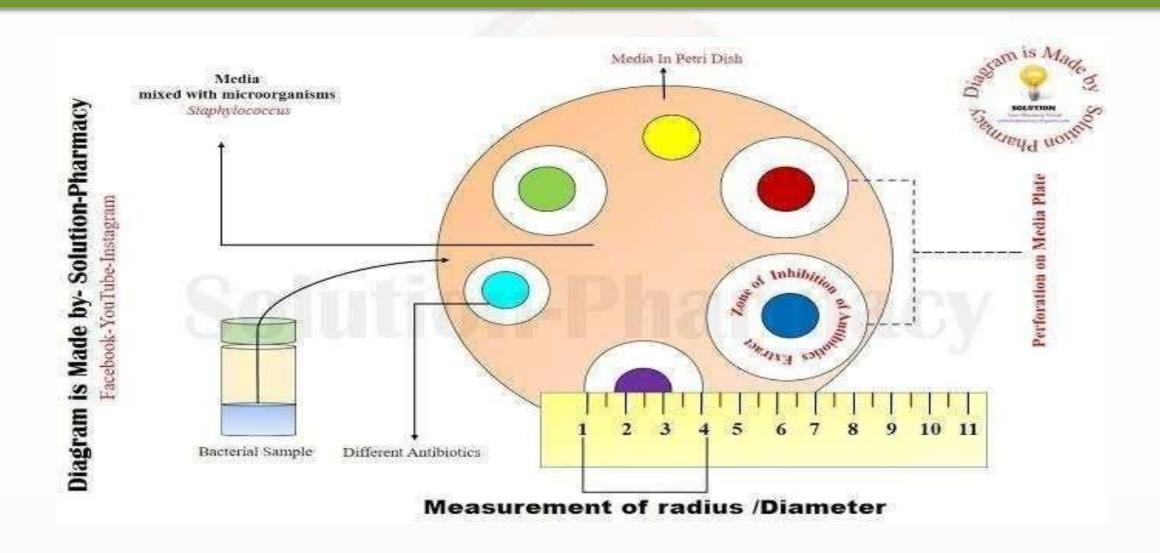
- A previously liquefied medium with the required quantity of microbial suspension is inoculated
- The suspension is added to the medium at a temperature between 40-50 degree and inoculated medium is immediately poured
- The solution are applied to the surface of the solider medium in sterile cylinder or in ager cavities
- They are incubated for about 18 hours at the temperature indicated

• The quantities estimation of antibiotic is done by accurately measuring the diameter or areas of the circular inhibition zones.

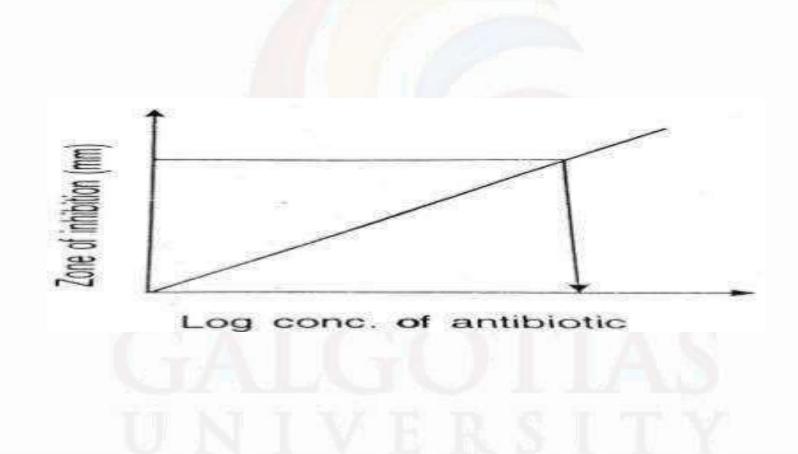
Simple image



Measurement of zone of inhibition

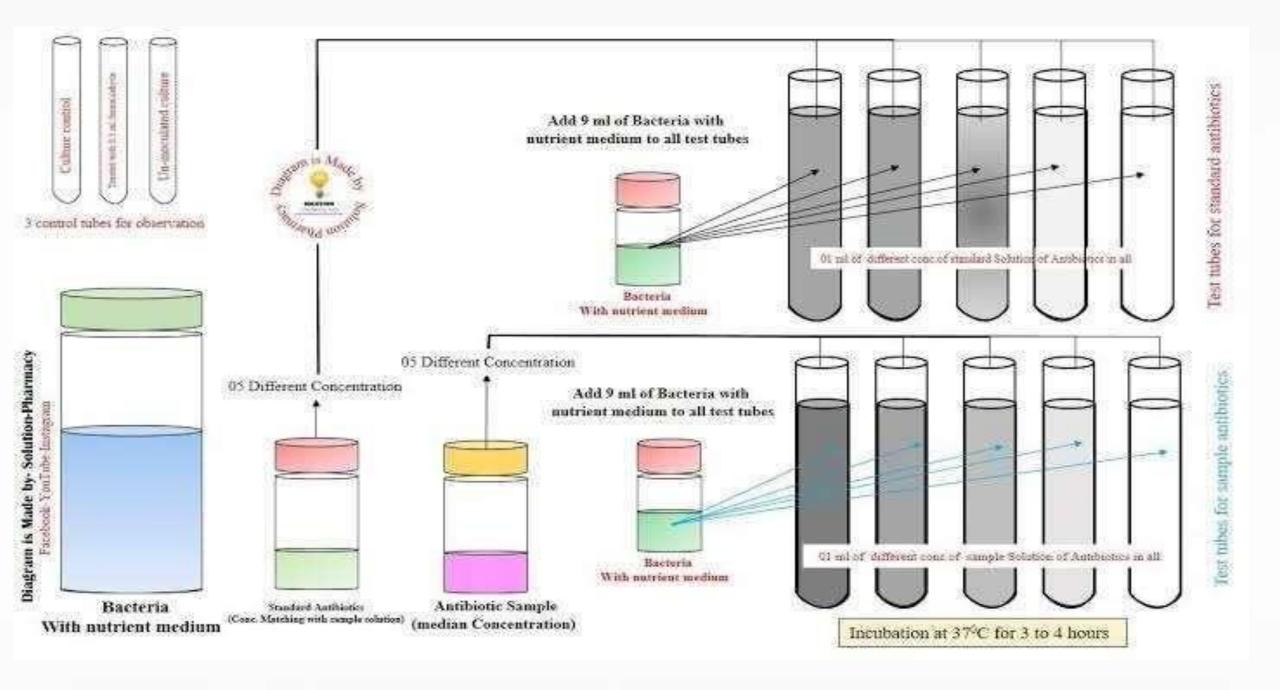


Standard curve of microbial assay of antibiotic (sample)



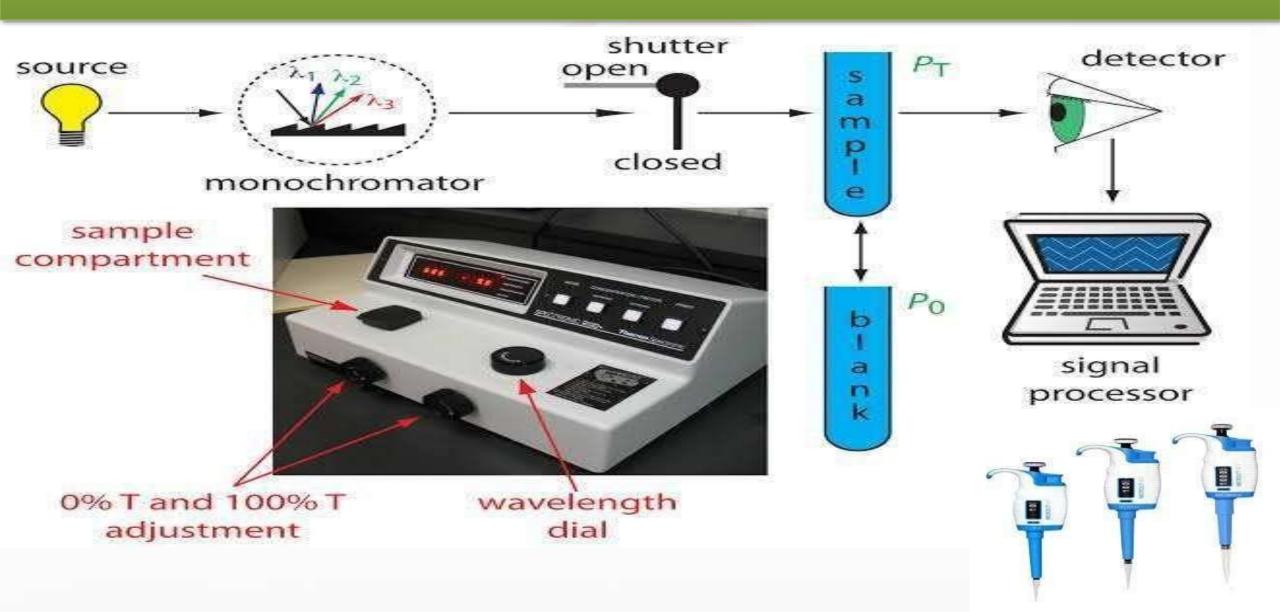
B. Turbidimetric or Tube Assay method

- Advantage- shorter incubation period for the growth of the test organism(usually 3 to 4 hrs)
- Disadvantage-The presence of solvent residues inhibitory substances affects more.
- Not recommended for cloudy or turbid preparation.
- Five different concentration of the standard solution are prepared for preparing the standard curve.
- 1mm of each concentration of the standard solution of the sample solution are placed in each of the tubes in duplicate at 9 ml of nutrients medium previously seeded with the appropriate test organism at to each other



- Five tubes containing the inoculated culture medium with standard drug with a specific dose and test organism.
- Five tubes containing culture medium with test organism and the test sample with different dosages.
- Another one treated immediately with 0.5 ml of dilute formaldehyde solution(blank)
- All the tubes are placed in an incubator and maintain at the specified temperature- 37°C for 3 to 4 hour.
- The growth of the test organism is measured by determining the absorbance at 530 nm of its against the blank.
- The standard calibration card is prepared and the absorbance obtained for the sample is plotted on it to obtain the concentration of the test antibiotic

SPECTROMETER FOR ABSORBANCE



Sample graph

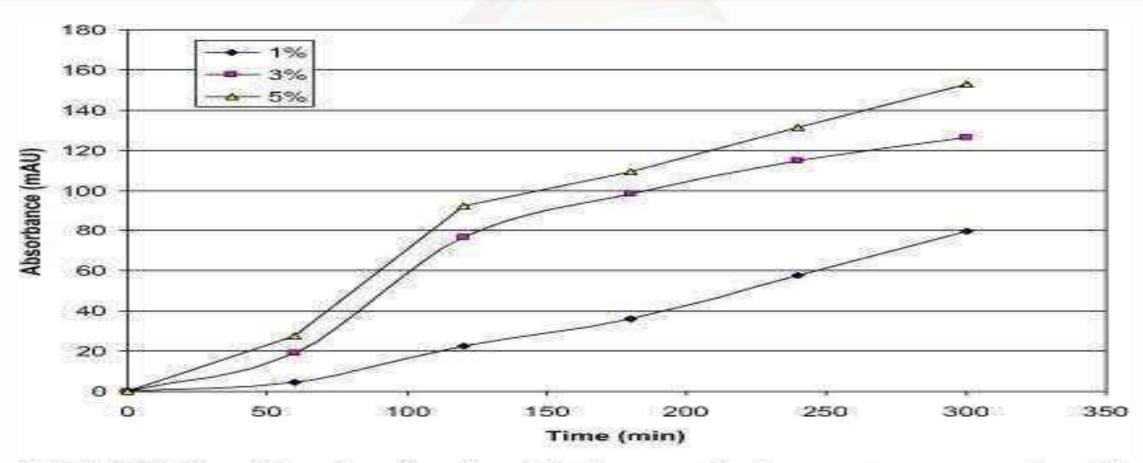


FIGURE 2 - Graph of microbial growth in tryptone soy broth inoculated with *Escherichia coli* (ATCC 8739) at 1%, 3% and 5% containing apramycin sulfate at 30 μg mL⁻¹.